

loss of bone due to immobilization. Nevertheless, calcium supplements *will* prevent the loss of bone due to inadequate calcium intake. Our mistake has been to generalize from studies in the immediate postmenopause, which is dominated by the readjustments produced by estrogen withdrawal, to periods either earlier or later, for which there is ample evidence that a high calcium intake is bone sparing.¹⁸⁻²⁰

Although it is never too late to start to assure an adequate calcium intake, it must also be acknowledged that the roots of the problem go deep. The building of an optimal bone mass, particularly during the teen years and young adulthood, is of critical importance. Many teenagers today have such low calcium intakes²¹ that there is no possibility that they can repeal the law of the conservation of mass and make much of a skeleton from the raw materials they provide their bodies. It is today's teenagers with low calcium intakes who are likely to be our hip fracture patients in 60-plus years from now. We are doomed to a perpetual game of catch-up until we can implement effective populationwide strategies to increase the level of calcium intake. This is an achievable goal, just as years ago ways were found to increase intakes of such trace nutrients as fluoride and iodine. If one is tempted to think about this as medicating the population, it may be instructive to bear two facts in mind: the primitive human calcium intake—the one, presumably, to which our physiology is adapted—is well in excess of 1,500 mg per day²²; and the natural food sources of our closest primate relatives provide them with diets containing roughly four times the calcium nutrient density of a typical first-world human diet.

Next, there is the importance of maintaining physical activity, both before and after a fracture. Our skeletons are, after all, mechanical systems, and the principal intrinsic stimulus to their self-maintenance is mechanical loading. Not only does physical activity help preserve bone mass, but it probably is an important factor in the maintenance of adequate remodeling, without which fatigue damage will accumulate. It is difficult to increase bone mass very much by increasing physical activity; unfortunately, it is easy to lose mass with inactivity. So maintaining load-bearing activity is critical.

Finally, a brief word about pharmacotherapy. Recent promising reports describing the use of bisphosphonates indicate that these agents may have a useful role in the management of patients who already have the disorder.^{23,24} Many years of experience have been accumulated with their use in Paget's disease, and so their long-term safety is reasonably assured. They are, however, remodeling suppressors, and their long-term effects in persons who already have compromised skeletal strength will have to await further study.

Fluoride is another issue. Riggs is cautious, which is understandable, given his recently reported experience in which fluoride did not decrease fracture incidence despite a nearly 40% increase in bone mass.¹¹ The protocol for his study, however, dictated not by the investigators but by the sponsoring agency, called both for what is now recognized to be a toxic dose of fluoride and for administration in a form that is known to produce excess gastrointestinal irritation.²⁵ The high peak blood concentrations produced by such therapy may well produce a degree of osteoblast toxicity that lower, but more sustained, blood levels do not. Certainly they will produce areas of hypercrystallinity in newly deposited bone mineral that will result in bone of different mechanical properties from bone mineralized out of a medium with a

lower fluoride concentration. Fluoride is now approved for the treatment of osteoporosis in at least eight European countries and should not yet be counted out of the running in the US.

ROBERT P. HEANEY, MD

John A. Creighton University Professor
Creighton University
Omaha, Nebraska

REFERENCES

1. Schneider EL, Guralnik JM: The aging of America—Impact on health care costs. *JAMA* 1990; 263:2335-2340
2. Kelsey JL: Risk factors for osteoporosis and associated fractures, *In* Proceedings of Special Topic Conference on Osteoporosis. Public Health Rep 1989; S104:14-20
3. Nagant de Deuxchaisnes C, Devogelaer JP: Increase in the incidence of hip fractures and of the ratio of trochanteric to cervical hip fractures in Belgium (Letter). *Calcif Tissue Int* 1988; 42:201-203
4. Finsen V, Benum P: Changing incidence of hip fractures in rural and urban areas of central Norway. *Clin Orthop* 1987; 218:104-110
5. Boyce WJ, Vessey MP: Rising incidence of fracture of the proximal femur. *Lancet* 1985; 1:150-151
6. Riggs BL: Overview of osteoporosis. *West J Med* 1991 Jan; 154:63-77
7. Heaney RP: Osteoporotic fracture space: An hypothesis. *Bone Miner* 1989; 6: 1-13
8. Mosekilde L: Age-related changes in vertebral trabecular bone architecture—Assessed by a new method. *Bone* 1988; 9:247-250
9. Eventov I, Frisch B, Cohen Z, Hammel I: Osteopenia, hematopoiesis and bone remodeling in iliac crest and femoral biopsies. *Bone* 1991, in press
10. Heaney RP, Avioli LV, Chesnut CH III, Lappe J, Recker RR, Brandenburger GH: Osteoporotic bone fragility—Detection by ultrasound transmission velocity. *JAMA* 1989; 261:2986-2990
11. Riggs BL, Hodgson SF, O'Fallon WM, et al: Effect of fluoride treatment on the fracture rate in postmenopausal women with osteoporosis. *N Engl J Med* 1990; 322:802-809
12. Delmi M, Rapin CH, Bengoa JM, Delmas PD, Vasey H, Bonjour JP: Dietary supplementation in elderly patients with fractured neck of the femur. *Lancet* 1990; 335:1013-1016
13. Holbrook TL, Barrett-Connor E, Wingard DL: Dietary calcium and risk of hip fractures: 14-year prospective population study. *Lancet* 1988; 2:1046-1049
14. Matkovic V, Kostial K, Simonovic I, Buzina R, Brodarec A, Nordin BEC: Bone status and fracture rates in two regions of Yugoslavia. *Am J Clin Nutr* 1979; 32:540-549
15. LaCroix AZ, Wienpahl J, White LR, et al: Thiazide diuretic agents and the incidence of hip fracture. *N Engl J Med* 1990; 322:286-290
16. NIH Consensus Conference: Osteoporosis. *JAMA* 1984; 252:799-802
17. Heaney RP: Estrogen-calcium interactions in the postmenopause: A quantitative description. *Bone Miner* 1990; 11:67-84
18. Dawson-Hughes B, Dallal GE, Krall EA, Sadowski L, Sahyoun N, Tannenbaum S: A controlled trial of the effect of calcium supplementation on bone density in postmenopausal women. *N Engl J Med* 1990; 323:878-883
19. Baran D, Sorensen A, Grimes J, et al: Dietary modification with dairy products for preventing vertebral bone loss in premenopausal women: A three-year prospective study. *J Clin Endocrinol Metab* 1990; 70:264-270
20. Cumming RG: Calcium intake and bone mass: A quantitative review of the evidence. *Calcif Tissue Int* 1990; 47:194-201
21. Carroll MD, Abraham S, Dresser CM: Dietary intake source data: United States, 1976-80. *Vital Health Stat* [11] 1983; 11:1-483
22. Eaton SB, Konner M: Paleolithic nutrition—A consideration of its nature and current implications. *N Engl J Med* 1985; 312:283-289
23. Storm T, Thamsborg G, Steiniche T, Genant JK, Sorensen OH: Effect of intermittent, cyclical etidronate therapy on bone mass and fracture rate in postmenopausal osteoporosis. *N Engl J Med* 1990; 322:1265-1271
24. Watts NB, Harris ST, Genant HK, et al: Intermittent cyclical etidronate treatment of postmenopausal osteoporosis. *N Engl J Med* 1990; 323:73-79
25. Heaney RP, Baylink DJ, Johnston CC Jr, et al: Fluoride therapy for vertebral crush fracture syndrome: Status report 1988. *Ann Intern Med* 1989; 111:678-680

Nitric Oxide, Nitrovasodilators, and L-Arginine—An Unusual Relationship

CERTAINLY ONE OF THE MOST interesting and potentially far-reaching discoveries that has taken shape over the past few years concerns a novel mammalian pathway that leads to the formation of nitric oxide from the amino acid L-arginine.¹ This unusual biochemical pathway brought together seemingly unrelated fields of research. Nitric oxide formation has now been found in a number of cell types, but the first and, until now, the most thorough characterizations have been in macrophages, endothelial cells, and cells of the central ner-

vous system. The action of nitric oxide generated through this pathway is dependent on the cell type that produces it. Endothelium-derived nitric oxide leads to vascular smooth muscle relaxation, and, although the debate continues, most of the evidence supports that nitric oxide is endothelium-derived relaxing factor (EDRF) and that it acts by activating the enzyme guanylate cyclase in smooth muscle tissue. In the central nervous system, nitric oxide acts in a manner similar to that from endothelial cells in that it signals an adjacent cell and in this case another neuron. Macrophage-derived nitric oxide is distinct in many ways from the other two cell types, and evidence supports a role for this nitric oxide in the cytostatic activity that is a critical function of these activated immune system cells.

The history of this pathway is relatively short and illustrative of the convergence of apparently disparate research areas. During the late 1970s, a number of key observations concerning guanylate cyclase were made whose overall importance and relationship to this pathway became clear later. In particular, Murad and colleagues showed that nitric oxide activates guanylate cyclase,² Craven and DeRubertis showed that this nitric oxide activation is dependent on heme,³ Ignarro and co-workers found that nitric oxide leads to the relaxation of smooth muscle,⁴ and Furchgott and Zawadzki showed that the endothelium was required for vascular smooth muscle relaxation.⁵ Also in the late 1970s, a number of laboratories involved with studies of the brain contributed to the growing idea that nitric oxide activated guanylate cyclase and that arginine and glutamate stimulated guanylate cyclase activity as well. In the early to middle 1980s, studies carried out with macrophages provided some key pieces to the puzzle. In experiments related to the endogenous formation of carcinogenic *N*-nitrosamines, Tannenbaum and co-workers showed that mammals were capable of nitrate synthesis.⁶ Continuing studies in that laboratory showed that nitrate urinary levels were significantly elevated after treating rats with immunostimulants such as lipopolysaccharide.⁷ Then in 1985 Stuehr and Marletta determined that macrophages treated with lipopolysaccharide synthesized nitrite and nitrate,⁸ and subsequent studies showed that lymphokines such as interferon gamma could also stimulate this synthesis.⁹ The stage was set in 1987 and 1988 for the convergence of the findings in the central nervous system (CNS), endothelium, and immune system. Iyengar and associates and Hibbs and colleagues showed that the precursor for nitrite and nitrate in macrophages was L-arginine,^{10,11} and, in addition, Iyengar and co-workers showed that the nitrite and nitrate were derived from the guanido group of arginine.¹⁰ Hibbs and colleagues also showed that arginine was required for macrophage-induced tumor cell cytostasis.¹² The chemical identity of EDRF had been the subject of much speculation and research, but it was Moncada and associates who provided the first definitive proof that endothelial cells were capable of nitric oxide synthesis.¹³ The solution instability of nitric oxide suggested that it was likely to be the intermediate that led to nitrite and nitrate in the macrophage studies mentioned above. Marletta and co-workers and Hibbs and associates provided direct proof that this was indeed the case.^{14,15} Moncada and colleagues then showed that the endothelial cell nitric oxide was derived from L-arginine,¹⁶ and Garthwaite and co-workers showed that nitric oxide was produced by glutamate stimulation of *N*-methyl-D-aspartate receptors.¹⁷ It was now clear that the L-arginine to

the nitric oxide pathway was the common theme in all of these observations.

An immediate outcome of these findings is that they provide a rational mechanism for the action of nitrovasodilators such as nitroglycerin. Nitroglycerin is known to decompose to nitric oxide under biologic conditions, and although the series of reactions is mechanistically complex and not completely understood, the reaction is facilitated in the presence of thiols such as glutathione. Therefore, nitroglycerin and related nitrovasodilators act by circumventing the enzymic generation of nitric oxide. It is particularly satisfying that the study of this pathway has led to an understanding of the mechanism of action of such an important class of drugs in use for more than 100 years and, perhaps more important, points the way toward the design of agents that could potentially mimic the pathway more successfully. As mentioned, there is not uniform agreement on the identity of EDRF. The debate has focused on the properties of EDRF generated from stimulated endothelial cells in a smooth muscle bioassay versus the action of nitric oxide and various *S*-nitroso compounds in the same bioassay. The answer will come from studies on the enzymology of the pathway in the various cell types, but at this point it appears that it is nitric oxide that is released by the enzyme. The solution decomposition of nitric oxide generates a number of products that readily nitrosate thiols and amines which can, especially in the case of *S*-nitroso compounds, cause smooth muscle relaxation.

The regulation and biochemistry of the pathway are two aspects under investigation at present. The results of these studies are likely to provide key pieces of information with regard to the potential for cell-type-specific drug interactions. Some important differences have already emerged where it is clear that the macrophage is distinct from the other cell types in a number of respects. Consistent with the function as a signaling agent, nitric oxide synthesis occurs in a burst in both endothelial and CNS cells. The activity in these cells requires calcium ion (Ca^{2+}), and a report of the enzyme purification from rat brain showed a requirement for Ca^{2+} -complexed calmodulin and a single polypeptide of 150,000 daltons.¹⁸ Because of the similarities in the CNS and endothelial cell activity, it may well turn out that the endothelial cell also shows a calmodulin requirement. In macrophages the activity cannot be detected in untreated cells, but after stimulation and subsequent protein synthesis, the cells continuously synthesize nitric oxide. The activity in macrophages is enhanced by divalent metals with magnesium ion being the most effective; these metals are not required, however. In addition, the cofactor tetrahydrobiopterin has also been identified as a requirement in macrophages.^{19,20} The most common role for this cofactor is in aromatic amino acid hydroxylation such as with phenylalanine hydroxylase. Experiments suggest that this is the function for this cofactor in macrophages. On the other hand, although there does not appear to be a low-molecular-weight cofactor involved in the CNS or endothelium, a relatively tightly bound pterin has not been ruled out. A number of arginine-related compounds have been shown to inhibit the reaction, most notably *N*^G-methyl-L-arginine, which inhibits the activity in all cells examined to date. L-Canavanine, however, appears to be a specific inhibitor for the macrophage and neutrophil, and, while not potent, it at least shows that selective inhibition is possible.

The stage is now set for exciting developments in the

application of this research to the development of new drugs. It is clear that substantial differences exist between the basic enzymology of the macrophage compared with that of the CNS and endothelial cells and in the regulation of the pathways in those cells. The extent to which these differences can be exploited will determine just how fruitful this area will be for drug development. It is difficult to see any advantage in the inhibition of the activity in endothelial cells, but inhibition of the activity in the macrophage may be useful in the treatment of endotoxemic shock, inflammatory bowel disease, and arthritis and in cytokine therapy. The extensive use of the nitrovasodilators in the treatment of angina and, as brought out in the article in this issue by Ignarro, Ross, and Tillisch,²¹ their potential in other cardiovascular diseases suggest that attention should be given to the development of new vasodilators with more controlled chemistry.

MICHAEL A. MARLETTA, PhD
Associate Professor of Medicinal Chemistry,
College of Pharmacy
Associate Professor of Biological Chemistry,
School of Medicine
University of Michigan
Ann Arbor, Michigan

REFERENCES

1. Marletta MA: Nitric oxide: Biosynthesis and biological significance. *Trends Biochem Sci* 1989; 14:488-492
2. Arnold WP, Mittal CK, Katsuki S, Murad F: Nitric oxide activates guanylate cyclase and increases guanosine 3':5'-cyclic monophosphate levels in various tissue preparations. *Proc Natl Acad Sci USA* 1977; 74:3203-3207
3. Craven PA, DeRubertis FR: Requirement for heme in the activation of purified guanylate cyclase by nitric oxide. *Biochim Biophys Acta* 1983; 745:310-321
4. Gruetter CA, Barry BK, McNamara DB, Gruetter DY, Kadowitz PJ, Ignarro L: Relaxation of bovine coronary artery and activation of coronary arterial guanylate cyclase by nitric oxide, nitroprusside and a carcinogenic nitrosoamine. *J Cyclic Nucleotide Res* 1979; 5:211-224
5. Furchgott RF, Zawadzki JV: The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature (Lond)* 1980; 288:373-376
6. Green LC, Ruiz de Luzuriaga K, Wagner DA, et al: Nitrate biosynthesis in man. *Proc Natl Acad Sci USA* 1981; 78:7764-7768
7. Wagner DA, Young VR, Tannenbaum S: Mammalian nitrate biosynthesis: Incorporation of $^{15}\text{NH}_3$ into nitrate is enhanced by endotoxin treatment. *Proc Natl Acad Sci USA* 1983; 80:4518-4531
8. Stuehr DJ, Marletta MA: Mammalian nitrate biosynthesis: Mouse macrophages produce nitrite and nitrate in response to *Escherichia coli* lipopolysaccharide. *Proc Natl Acad Sci USA* 1985; 82:7738-7742
9. Stuehr DJ, Marletta MA: Induction of nitrite/nitrate synthesis in murine macrophages by BCG infection, lymphokines, or interferon- γ . *J Immunol* 1987; 139:518-525
10. Iyengar R, Stuehr DJ, Marletta MA: Macrophage synthesis of nitrite, nitrate, and *N*-nitrosamines: Precursors and role of the respiratory burst. *Proc Natl Acad Sci USA* 1987; 84:6369-6373
11. Hibbs JB Jr, Taintor RR, Vavrin Z: Macrophage cytotoxicity: Role for L-arginine deiminase and imino nitrogen oxidation to nitrite. *Science* 1987; 235:473-476
12. Hibbs JB Jr, Vavrin Z, Taintor RR: L-Arginine is required for expression of the activated macrophage effector mechanism causing selective metabolic inhibition in target cells. *J Immunol* 1987; 138:550-565
13. Palmer RMJ, Ferrige AG, Moncada S: Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature* 1987; 327:524-526
14. Marletta MA, Yoon PS, Iyengar R, Leaf CD, Wishnok JS: Macrophage oxidation of L-arginine to nitrite and nitrate: Nitric oxide is an intermediate. *Biochemistry* 1988; 27:8706-8711
15. Hibbs JB, Taintor RR, Vavrin Z, Rachlin EM: Nitric oxide: A cytotoxic activated macrophage effector molecule. *Biochem Biophys Res Commun* 1988; 157:87-94 (erratum 158:624)
16. Palmer RMJ, Ashton DS, Moncada S: Vascular endothelial cells synthesize nitric oxide from L-arginine. *Nature* 1988; 333:664-666
17. Garthwaite J, Charles SL, Chess-Williams R: Endothelium-derived relaxing factor release on activation of NMDA receptors suggests a role as intercellular messenger in the brain. *Nature* 1988; 336:385-388
18. Bredt DS, Snyder SH: Isolation of nitric oxide synthetase, a calmodulin-requiring enzyme. *Proc Natl Acad Sci USA* 1990; 87:682-685
19. Tayeh MA, Marletta MA: Macrophage oxidation of L-arginine to nitric oxide, nitrite, and nitrate—Tetrahydrobiopterin is required as a cofactor. *J Biol Chem* 1989; 264:19654-19658
20. Kwon NS, Nathan CF, Stuehr DJ: Reduced biopterin as a cofactor in the generation of nitrogen oxides by murine macrophages. *J Biol Chem* 1989; 264:20496-20501
21. Ignarro LJ, Ross G, Tillisch J: Pharmacology of endothelium-derived nitric oxide and nitrovasodilators. *West J Med* 1991; 154:51-62